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<p>(54) Title: CELL CONCENTRATION AND LYSATE CLEARANCE USING PARAMAGNETIC PARTICLES</p> <p>(57) Abstract</p> <p>Methods are disclosed for using paramagnetic particles to concentrate or harvest cells. Methods are also disclosed for clearing a solution of disrupted biological material, such as a lysate of cells or a homogenate of mammalian tissue. Methods are also disclosed for using paramagnetic particles to isolate target nucleic acids, such as RNA or DNA, from a solution cleared of disrupted biological material using the same type or a different type of paramagnetic particle. Kits are also disclosed for use with the various methods of the present invention. Nucleic acids isolated according to the present methods and using the present kits are suitable for immediate use in downstream processing, without further purification.</p>		

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CELL CONCENTRATION AND LYSATE CLEARANCE USING PARAMAGNETIC PARTICLES

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CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Application Number 60/134,156, filed May 14, 1999.

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STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

Not applicable.

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TECHNICAL FIELD

This invention relates generally to the use of magnetically responsive particles, such as magnetically responsive silica gel particles or magnetically responsive ion exchange particles, to harvest or to concentrate cells or biological tissue. This invention also relates to the use of such particles to clear lysates or homogenates of such cells or tissue. This invention relates, furthermore, to the use of such particles to isolate target nucleic acids, such as plasmid DNA, chromosomal DNA, DNA fragments, total RNA, mRNA, or RNA/DNA hybrids from non-target material in a cell lysate.

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BACKGROUND OF THE INVENTION

Cells in a liquid culture must be concentrated or harvested before they can be preserved for later use, stained for direct analysis, or processed to isolate target specific materials therefrom. Most cell harvesting and concentration techniques involve centrifugation, filtration, or a combination of centrifugation and filtration. (See, e.g. Molecular Cloning, (1989) ed. by Sambrook et al., pp 2.22 and filtration system reference). Unfortunately, neither filtration nor centrifugation is amenable to automation. Specifically, neither can be performed at basic pipettor-diluter robotics stations, such as the Biomec[®]. When it becomes necessary to isolate or analyze certain types of material in the interior of a cell, such as a target nucleic acid or a protein, the cell membrane must be disrupted and the contents of the cell released into the solution surrounding the cell. Such disruption can be accomplished by mechanical means (e.g., by sonication or by blending in a mixer), by

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enzymatic digestion (e.g. by digestion with proteases), or by chemical means (e.g., by alkaline lysis followed by addition of a neutralization solution). Whatever means is used to disrupt a cell, the end product, referred to herein as a lysate solution, consists of the target material and many contaminants, including cell debris. The lysate solution must be cleared of as many of the large contaminants as possible before the target material can be further isolated therefrom. Either or both of the same two means described above, i.e. centrifugation and filtration, have been used to clear lysate solutions prior to further processing. However, for reasons given above, neither means of clearing a lysate solution is amenable to automation.

Many different systems of materials and methods have been developed for use in the isolation of nucleic acids from cleared lysate solutions. Many such systems are silica based, such as those which employ controlled pore glass, filters embedded with silica particles, silica gel particles, resins comprising silica in the form of diatomaceous earth, glass fibers or mixtures of the above. Each such silica-based solid phase separation system is configured to reversibly bind nucleic acid materials when placed in contact with a medium containing such materials in the presence of chaotropic agents. The silica-based solid phases are designed to remain bound to the nucleic acid material while the solid phase is exposed to an external force such as centrifugation or vacuum filtration to separate the matrix and nucleic acid material bound thereto from the remaining media components. The nucleic acid material is then eluted from the solid phase by exposing the solid phase to an elution solution, such as water or an elution buffer. Numerous commercial sources offer silica-based resins designed for use in centrifugation and/or filtration isolation systems, e.g. Wizard[®] DNA purification systems products from Promega Corporation (Madison, Wisconsin, U.S.A.), or the QiaPrep[®] DNA isolation systems from Qiagen Corp. (Chatsworth, California, U.S.A.). Unfortunately, the type of silica-based solid phases described above all require one use centrifugation or filtration to perform the various isolation steps in each method, limiting the utility of such solid phases in automated systems.

Magnetically responsive solid phases, such as paramagnetic or superparamagnetic particles, offer an advantage not offered by any of the silica-based solid phases described above. Such particles could be separated from a solution by turning on and off a magnetic force field, or by moving a container on to and off of a magnetic separator. Such activities would be readily adaptable to automation.

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Magnetically responsive particles have been developed for use in the isolation of nucleic acids. Such particles generally fall into either of two categories, those designed to reversibly bind nucleic acid materials directly, and those designed to reversibly bind nucleic acid materials through an intermediary. For an example of particles of the first type, see
5 silica based porous particles designed to reversibly bind directly to DNA, such as MagneSil™ particles from Promega, or BioMag® magnetic particles from PerSeptive Biosystems. For examples of particles and systems of the second type designed to reversibly bind one particular type of nucleic acid (mRNA), see the PolyATract® Series 9600™ mRNA Isolation System from Promega Corporation (Madison, Wisconsin, U.S.A.); or the
10 streptavidin coated microsphere particles from Bangs Laboratories (Carmel, Indiana, U.S.A.). Both of these systems employ magnetically responsive particles with streptavidin subunits covalently attached thereto, and biotin with an oligo(dT) moiety covalently attached thereto. The biotin-oligo(dT) molecules act as intermediaries, hybridizing to the poly(A) tail of mRNA molecules when placed into contact therewith, then binding to the streptavidin on
15 the particles. The mRNA molecules are then released in water.

Indirect binding magnetic separation systems for nucleic acid isolation or separation require at least three components, i.e. magnetic particles, an intermediary, and a medium containing the nucleic acid material of interest. The intermediary/nucleic acid hybridization reaction and intermediary/particle binding reaction often require different solution and/or
20 temperature reaction conditions from one another. Each additional component or solution used in the nucleic acid isolation procedure adds to the risk of contamination of the isolated end product by nucleases, metals, and other deleterious substances.

Various types of magnetically responsive silica based particles have been developed for use as solid phases in direct or indirect nucleic acid binding isolation methods. One such
25 particle type is a magnetically responsive glass bead, preferably of a controlled pore size. See, e.g. Magnetic Porous Glass (MPG) particles from CPG, Inc. (Lincoln Park, New Jersey, U.S.A.); or porous magnetic glass particles described in U.S. Pat. No.'s 4,395,271; 4,233,169; or 4,297,337. Nucleic acid material tends to bind very tightly to glass, however, so that it can be difficult to remove once bound thereto. Therefore, elution efficiencies from
30 magnetic glass particles tend to be low compared to elution efficiencies from particles containing lower amounts of a nucleic acid binding material such as silica.

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Another type of magnetically responsive particle designed for use as a solid phase in direct binding and isolation of nucleic acids, particularly DNA, is a particle comprised of agarose embedded with smaller ferromagnetic particles and coated with glass, e.g. U.S. Patent 5,395,498. Yet another type of magnetically responsive particle designed for direct
5 binding and isolation of nucleic acids is produced by incorporating magnetic materials into the matrix of polymeric silicon dioxide compounds, e.g. German Patent Application No. DE 43 07 262. The latter two types of magnetic particles, the agarose particle and the polymeric silicon dioxide matrix, tend to leach iron into a medium under the conditions required to bind nucleic acid materials directly to each such magnetic particle. It is also difficult to
10 produce such particles with a sufficiently uniform and concentrated magnetic capacity to ensure rapid and efficient isolation of nucleic acid materials bound thereto.

Magnetically responsive beads designed for use in the isolation of target polymers, such as nucleic acids, and methods for their use therein are described in U.S. Pat. No. 5,681,946 and in International Publication No. WO 91/12079. These last beads are designed
15 to become nonspecifically associated with the target polymer, only after the target polymer is precipitated out of a solution comprising the target polymer and the beads. Magnetic force is used to isolate the beads and polymer associated therewith from the solution. The magnetically responsive beads recommended for use in this last system are "finely divided magnetizable material encapsulated in organic polymer." ('946 Patent, col. 2, line 53).

20 A variety of solid phases have also been developed with ion exchange ligands capable of exchanging with nucleic acids. However, such systems are generally designed for use as a solid phase of a liquid chromatography system, for use in a filtration system, or for use with centrifugation to separate the solid phase from various solutions. Such systems range in complexity from a single species of ligand covalently attached to the surface of a
25 filter, as in DEAE modified filters (e.g., CONCERT[®] isolation system, Life Technology Inc., Gaithersburg, MD, U.S.A.), to a column containing two different solid phases separated by a porous divider (e.g., U.S. Patent No. 5,660,984), to a chromatography resin with pH dependent ionizable ligands covalently attached thereto (e.g., U. S. Pat. No. 5,652,348).

30 Materials and methods are needed which enable one to automate as many steps as possible to quickly and efficiently isolate target nucleic acids from cells or mammalian tissue. Specifically, methods and materials are needed for the concentration or harvesting of cells, for the clearing of solutions of disrupted cells or tissue, and for the isolation of target

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nucleic acids from such cleared solutions, wherein labor-intensive steps such as filtration or centrifugation are not required. The present invention addresses each of these needs. Nucleic acids isolated according to the present method can be used in a variety of applications, including restriction digestion and sequencing.

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BRIEF SUMMARY OF THE INVENTION

In the methods of the present invention, magnetic particles are used to process biological material. In one embodiment, the present invention is a method of concentrating or harvesting cells comprising the steps of: (a) combining a solution with cells contained
10 therein, such as an overnight culture of bacteria in a growth medium or white cells in whole blood with magnetic particles under conditions wherein the cells form a complex with the magnetic particles; and (b) isolating the magnetic particle/cell complex from the solution by application of magnetic force, e.g., by means of a magnet.

In another embodiment, the present invention is a method of clearing disrupted
15 biological material, such as a cell lysate or a homogenate of mammalian tissue, comprising the steps of: (a) providing a solution comprising a disrupted biological material, such as a cell lysate or homogenized tissue; (b) combining the solution with magnetic particles under conditions wherein the disrupted biological material forms a complex with the magnetic particles; and (c) isolating the complex from the solution by application of magnetic force.

20 In yet another embodiment, the present invention is a method of isolating a target nucleic acid from a solution of disrupted biological material, comprising the target nucleic acid, a first non-target material, and a second non-target material, comprising the steps of: (a) combining a solution of the disrupted biological material with first magnetic particles under conditions wherein the first non-target material forms a first complex with the first
25 magnetic particles; (b) separating the first complex from the solution of disrupted biological material by application of magnetic force, forming a cleared solution comprising the target nucleic acid and the second non-target material; (c) combining the cleared solution with second magnetic particles under conditions wherein the target nucleic acid adsorbs to the second magnetic particles, forming a second complex; (d) isolating the second complex from
30 the cleared solution; (e) washing the second complex by combining the second complex with a wash solution and separating the second complex from the wash solution by magnetic

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force; and (f) combining the washed second complex with an elution solution, under conditions wherein the target material is desorbed from the second magnetic particles.

In yet another embodiment, the present invention also consists of kits with at least one type of magnetic particle and at least one solution needed to practice one or more of the methods of the invention, described above. In one such embodiment, the present invention is a kit comprising: (a) a first container of first magnetic particles with the capacity to form a first complex with first non-target material in a first solution of disrupted biological material comprising the first non-target material and the target nucleic acid; and (b) a second container of second magnetic particles with the capacity to form a second complex with the target nucleic acid, under solution conditions designed to promote the specific adsorption of the target nucleic acid to the second magnetic particles.

The methods and materials of the present invention can be used to isolate target nucleic acids including, but not limited to plasmid DNA, total RNA, mRNA, RNA/DNA hybrids, amplified nucleic acids, and genomic DNA from a variety of contaminants, including but not limited to agarose and components of a bacteria, animal tissue, blood cells, and non-target nucleic acids. Applications of the methods and compositions of the present invention to isolate nucleic acids from a variety of different media will become apparent from the detailed description of the invention below. Those skilled in the art of this invention will appreciate that the detailed description of the invention is meant to be exemplary only and should not be viewed as limiting the scope of the invention.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a photograph of samples of plasmid DNA isolated with MagneSil™ particles (Promega) or varying amounts of MagIE-glycidyl-histidine particles, fractionated by gel electrophoresis and visualized by staining with ethidium bromide, as described in Example 6.

Figure 2 is a photograph of samples of plasmid DNA isolated from varying amounts of a culture of transformants of *E.coli* DH5α cells using centrifugation ("Spin") on MagneSil™ particles (Promega Corp.) ("Mag"), followed by fractionation by gel electrophoresis on a short run gel, and visualization by staining with ethidium bromide, as described in Example 7.

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Figure 3 is a photograph of the same gel shown in Figure 2, shot after electrophoresis was continued for a longer period of time.

Figure 4 is a photograph of samples of DNA and RNA isolated from a mouse liver homogenate, using MagIE-glycidyl-histidine particles, as described in Example 9, after
5 fractionation by gel electrophoresis and visualization by staining with ethidium bromide.

Figure 5 is a photograph of samples of DNA and RNA isolated from mouse spleen (lanes 2-5) and kidney (lanes 7-9), using MagIE-glycidyl-histidine particles, as described in Example 9, after the samples were fractionated by gel electrophoresis and visualized by staining with ethidium bromide, as described in Example 9.

10 Figure 6 is a photograph of mouse liver RNA and DNA, after digestion with DNase, fractionation by gel electrophoresis, and visualization by staining with ethidium bromide.

DETAILED DESCRIPTION OF THE INVENTION

The present invention will now be described in detail, in part, by reference to the
15 following definitions.

The term "solid phase" is used herein in a standard chromatographic sense, to refer to an insoluble, usually rigid, matrix or stationary phase which interacts with a solute, in this case a tissue or cell or target nucleic acid, in a solute mixture. In the methods and kits of the present invention magnetic particles function as a solid phase when added to various solute
20 mixtures.

The term "surface", as used herein, refers to the portion of the support material of a solid phase which comes into direct contact with a solution when the solid phase is combined therewith.

The term "silica gel" as used herein refers to chromatography grade silica gel, a
25 substance which is commercially available from a number of different sources. Silica gel is most commonly prepared by acidifying a solution containing silicate, e.g. by acidifying sodium silicate to a pH of less than 11, and then allowing the acidified solution to gel. See, e.g. silica preparation discussion in Kurt-Othmer Encyclopedia of Chemical Technology, Vol. 21, 4th ed., Mary Howe-Grant, ed., John Wiley & Sons, pub., 1997, p. 1021.

30 As used herein, the term "silica magnetic particles" refers to silica based solid phases which are further comprised of materials which have no magnetic field but which form a magnetic dipole when exposed to a magnetic field, i.e., materials capable of being

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magnetized in the presence of a magnetic field but which are not themselves magnetic in the absence of such a field.

The term "magnetic", as used herein refers to temporarily magnetic materials, such as ferrimagnetic or ferromagnetic materials. The term encompasses paramagnetic and
5 superparamagnetic materials.

The term "magnetic particle" refers to a matrix comprising a core of paramagnetic or superparamagnetic materials and a solid phase capable of forming a complex with a solute of interest.

The term "silica magnetic particles", as used herein refers to paramagnetic particles
10 comprising a superparamagnetic core coated with siliceous oxide, having a hydrous siliceous oxide adsorptive surface (i.e. a surface characterized by the presence of silanol groups).

The term "magnetic ion exchange particles", as used herein, refers to paramagnetic particles with ion exchange ligands covalently attached thereto.

The term "pH dependent ion exchange magnetic particles", as used herein, refers to
15 magnetic particles with a plurality of ion exchange ligands covalently attached thereto, which can act as cation exchangers at one pH and as anion exchangers at another pH. Such magnetic particles are particularly well suited for use in the methods and kits of the present invention, as their binding capacity to different substrates can be adjusted merely by varying the pH or salt conditions in a solution.

20 The term "pH dependent ion exchange silica magnetic particles", as used herein, refers to silica magnetic particles with a plurality of ion exchange ligands covalently attached thereto, which can act as cation exchangers at one pH and as anion exchangers at another pH. Such magnetic particles are particularly well suited for use in the methods and kits of the present invention, because substrates can selectively adsorb to the hydrous
25 siliceous oxide adsorptive surface of the particle through hydrophobic interactions, to the ion exchange ligands through ion exchange, or to both the surface and ion exchange ligands, depending upon solution conditions.

The term "nucleic acid" as used herein refers to any DNA or RNA molecule or a DNA/RNA hybrid molecule. The term includes plasmid DNA, amplified DNA or RNA
30 fragments, total RNA, mRNA, genomic DNA, and chromosomal DNA.

The term "target nucleic acid" as used herein refers to any particular species of nucleic acid to be isolated using magnetic particles according to a method of the present

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invention. The target nucleic acid is preferably at least 20 nucleotides long, more preferably at least 100 nucleotides long, and most preferably at least 1,000 nucleotides long.

The methods and kits of the present invention can be used to harvest or concentrate cells, to clear a solution of disrupted biological material, and/or to isolate a target nucleic acid from a solution, preferably from a solution of cleared disrupted biological material. In at least one step of each such method, a complex is formed in a solution between a solute and magnetic particles. The resulting complex is then isolated from or removed from the solution by the application of magnetic force. Magnetic particles suitable for use in any given step of the methods and kits of the present invention have the capacity to form a complex with the solute of interest in that particular step of the method.

The solute is the type of material to be isolated from or removed from a solution, using magnetic particles, according to a method of the present invention. Cells to be concentrated or harvested are the solute in the harvesting method of the present invention. Disrupted biological material is the solute in the lysate or homogenate clearing method of the invention. A target nucleic acid is the solute when magnetic particles are used to isolate the target nucleic acid from any solution comprising the target nucleic acid and other material, such as a cleared lysate or homogenate solution.

In one aspect of the methods of the present invention, cells are harvested or concentrated using magnetic particles which can form a complex with the cells, under solution conditions designed to promote the formation of the complex. Silica magnetic particles and pH dependent ion exchange magnetic particles are both suitable for use in harvesting or concentrating cells according to the method of the present invention. However, one of ordinary skill in the art could readily select other suitable magnetic particles for use in this particular embodiment of the invention.

Conditions which promote the formation of a magnetic particle/solute complex vary, depending upon the nature of the solute and on the characteristics of the solid phase component of the magnetic particle. For example, when the magnetic particles are ion exchange magnetic particles or pH dependent ion exchange particles, the complex is preferably formed as a result of ion exchange between the solute and ion exchange ligands at the surface of the particles. In order to promote such ion exchange interaction, there must be at least some salt present in the solution to promote ion exchange with the solute, and the pH of the solution must be within the range wherein the ion exchange ligand has a charge

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appropriate to exchange with the solute. When the magnetic particles are silica magnetic particles, the complex is preferably formed as a result of hydrophobic interactions between the solute and particles. When the magnetic particles are pH dependent ion exchange silica magnetic particles, the complex can be formed as a result of hydrophobic interactions
5 between the solute and the siliceous oxide surface of the particles, as a result of ion exchange between the solute and the ion exchange ligands, or as a result of a combination of the two types of interactions. Preferred salt, pH, and other solution conditions to be used to promote formation of a complex with any given preferred substrate isolated according to the present methods or using the present kits are described below.

10 When the solute is intact cells, the complex is preferably formed in the presence of a low molecular weight alcohol, such as ethanol or isopropanol.

When the solute is disrupted biological material, such as one finds in a cell lysate or tissue homogenate, and the magnetic particles are silica-based particles, the magnetic particle/solute complex is preferably formed in a solution which does not contain any more
15 than trace amounts of alcohol or of chaotropic salts. Both alcohol and chaotropic salts, such as guanidine thiocyanate or guanidine isothiocyanate, promote adsorption of nucleic acid materials to such particles. It is contemplated, however, that one could practice the present method of cell lysate clearance in the presence of alcohol or chaotropic salts if the concentration of magnetic particles in a homogenate or lysate solution were low enough to
20 clear the solution, but not high enough to adhere to a significant amount of the target nucleic acid in the solution.

When the solute is a target nucleic acid, formation of the complex is preferably done in the presence of at least one agent known to promote reversible adsorption of the target nucleic acid to the magnetic particles. The reversible adsorption reaction is preferably done
25 through specific adsorption between the target nucleic acid and magnetic particles, leaving non-target material in solution. For example, when the target nucleic acid is plasmid DNA being isolated from a cleared lysate solution, the plasmid DNA is combined with magnetic particles under conditions wherein the plasmid DNA forms a complex therewith while non-target materials, such as proteins, lipids, and chromosomal DNA remain in solution. When
30 the magnetic particle is an ion exchange magnetic particle, the complex is formed in the presence of a counterion and in a solution with a pH at which the ion exchange ligands have the capacity to exchange with the target nucleic acid. When the magnetic particles are silica

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magnetic particles, formation of the complex is preferably done in the presence of an agent selected from the group consisting of a low molecular weight alcohol, a high concentration of a non-chaotropic salt, and a chaotropic salt, or a combination of any of the above. For methods of adsorption and desorption of target nucleic acids to silica magnetic particles, which are suitable for use in the present invention, see international patent application number PCT/US98/01149 for METHODS OF ISOLATING BIOLOGICAL TARGET MATERIALS USING SILICA MAGNETIC PARTICLES, published as WO 98/31840, incorporated by reference herein.

The solid phase of the magnetic particles used in the present methods can be made of any common support material, including soft gel supports such as agarose, polyacrylamide, or cellulose, or hard support material such as polystyrene, latex, methacrylate, or silica.

When the solid phase support material is silica, it is preferably in the form of silica gel, siliceous oxide, solid silica such as glass or diatomaceous earth, or a mixture of two or more of the above. Silica based solid phases suitable for use in the pH dependent ion exchange matrixes of the present invention include the mixture of silica gel and glass described in U.S. Pat No. 5,658,548, the silica magnetic particles described in PCT Publication Number WO 98/31840, and solid phases sold by Promega Corporation for use in plasmid DNA isolation, i.e. Wizard[®] Minipreps DNA Purification Resin. Silica gel particles are particularly preferred for use as the solid phase in the pH dependent ion exchange matrix and methods of the present invention. Silica gel particles are stable at much higher pressures than solid phases made from soft gel support material, making the silica gel solid phases suitable for HPLC as well as LC and batch separation applications.

Silica magnetic particles can be used to concentrate cells, clear lysates, or isolate target nucleic acids according to the methods the present invention. When silica magnetic particles are employed, the silica-based surface material of the particle specifically interacts with the various solutes isolated or removed therewith.

When the silica magnetic particles have ion exchange ligands covalently attached thereto, the silica-based surface material acts primarily as a solid support for the ion exchange ligands, which enable the particles to form complexes with the various solutes to be isolated or removed from any given solution. When used to isolate a target nucleic acid, the ion exchange ligands are preferably capable of forming a complex with the target nucleic acid by exchanging therewith at one pH, and of releasing the target nucleic acid at another

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pH. The most preferred ion exchange ligands are ones which complex with the target nucleic acid at a pH which is lower than a neutral pH, and which release the target nucleic acid at about a neutral pH and in low salt conditions, so the target nucleic acid released therein can be used immediately, without concentration or further isolation. Such preferred ion exchange ligands and pH dependent ion exchange matrices which incorporate such ligands are described in U.S. Patent Application Ser. No. 09/312,172, for an invention titled pH DEPENDENT ION EXCHANGE MATRIX AND METHOD OF USE IN THE ISOLATION OF NUCLEIC ACIDS, incorporated by reference herein, an application filed concurrently with the provisional patent application on which the present non-provisional patent application is based.

When the solid support component of the pH dependent ion exchange matrix is a silica magnetic particle, the size of the particle is preferably selected as follows. Smaller silica magnetic particles provide more surface area (on a per weight unit basis) for covalent attachment to the plurality of ion exchange ligands, but smaller particles are limited in the amount of magnetic material which can be incorporated into such particles compared to larger particles. The median particle size of the silica magnetic particles used in a particularly preferred embodiment of the present invention is about 1 to 15 μm , more preferably about 3 to 10 μm , and most preferably about 4 to 7 μm . The particle size distribution may also be varied. However, a relatively narrow monodal particle size distribution is preferred. The monodal particle size distribution is preferably such that about 80% by weight of the particles are within a 10 μm range of the median particle size, more preferably within an 8 μm range, and most preferably within a 6 μm range.

The magnetic particles of the present invention can be porous or non-porous. When the magnetic particles are porous, the pores are preferably of a controlled size range sufficiently large to admit the target nucleic acid material into the interior of the solid phase particle, and to bind to functional groups or silica on the interior surface of the pores. When the magnetic particles are porous silica magnetic particles, the total pore volume of each silica magnetic particle, as measured by nitrogen BET method, is preferably at least about 0.2 ml/g of particle mass. The total pore volume of porous silica magnetic particles particularly preferred for use as components of the pH dependent ion exchange matrix of the

present invention, as measured by nitrogen BET, is preferably at least about 50% of the pore volume is contained in pores having a diameter of 600 Å or greater.

Silica magnetic particles may contain substances, such as transition metals or volatile organics, which could adversely affect the utility of target nucleic acids substantially contaminated with such substances. Specifically, such contaminants could affect downstream processing, analysis, and/or use of the such materials, for example, by inhibiting enzyme activity or nicking or degrading the target nucleic acids isolated therewith. Any such substances present in the silica magnetic particles used in the present invention are preferably present in a form which does not readily leach out of the particle and into the isolated biological target material produced according to the methods of the present invention. Iron is one such undesirable at least one contaminant, particularly when the biological target material is a target nucleic acid.

Iron, in the form of magnetite, is present at the core of particularly preferred forms of silica magnetic particles used as the solid phase component of the pH dependent ion exchange matrixes of the present invention. Iron has a broad absorption peak between 260 and 270 nanometers (nm). Target nucleic acids have a peak absorption at about 260 nm, so iron contamination in a target nucleic acid sample can adversely affect the accuracy of the results of quantitative spectrophotometric analysis of such samples. Any iron containing silica magnetic particles used to isolate target nucleic acids using the present invention preferably do not produce isolated target nucleic acid material sufficiently contaminated with iron for the iron to interfere with spectrophotometric analysis of the material at or around 260 nm.

The most preferred silica magnetic particles used in the matrixes and methods of the present invention, siliceous oxide coated silica magnetic particles, leach no more than 50 ppm, more preferably no more than 10 ppm, and most preferably no more than 5 ppm of transition metals when assayed as follows. Specifically, the particles are assayed as follows: 0.33 g of the particles (oven dried at 110°C) are combined with 20 ml. of 1N HCl aqueous solution (using deionized water). The resulting mixture is then agitated only to disperse the particles. After about 15 minutes total contact time, a portion of the liquid from the mixture is then analyzed for metals content. Any conventional elemental analysis technique may be employed to quantify the amount of transition metal in the resulting liquid, but inductively coupled plasma spectroscopy (ICP) is preferred.

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At least two commercial silica magnetic particles are particularly preferred for use in the present invention, BioMag[®] Magnetic Particles from PerSeptive Biosystems, and the MagneSil[™] Particles available from Promega Corporation (Madison, Wisconsin). Any source of magnetic force sufficiently strong to separate the silica magnetic particles from a solution would be suitable for use in the nucleic acid isolation methods of the present invention. However, the magnetic force is preferably provided in the form of a magnetic separation stand, such as one of the MagneSphere[®] Technology Magnetic Separation Stands (cat. no.'s Z5331 to 3, or Z5341 to 3) from Promega Corporation.

When magnetic particles are used to both clear a solution of disrupted biological material and to isolate a target nucleic acid therefrom, one can use the same type of particles or a different type of particles for clearing and isolation. For purposes of this disclosure, and to emphasize the flexibility in the invention, the particles used to clear the solution of disrupted biological material are referred to as first magnetic particles, while the particles used to isolate the target nucleic acid are referred to as second magnetic particles.

When the target nucleic acid is plasmid DNA, the second magnetic particles can be added directly to cleared lysate of bacteria transformed with the plasmid DNA, wherein the lysate is formed by alkaline lysis followed by clearance using first magnetic particles as described above. Alkaline lysis procedures suitable for use in the present invention can be found in Sambrook et al, Molecular Cloning, Vol. 1, 2nd ed. (pub. 1989 by Cold Spring Harbor Laboratory Press), pp. 1.25-1.28, and in Technical Bulletin No's 202, 225, and 259 (Promega Corp.). When the second silica magnetic particle is a pH dependent ion exchange particle, plasmid DNA from a lysate solution prepared as described above will form a complex with the pH dependent ion exchange particles upon combination therewith, provided the overall charge of the matrix is positive, and provided the charge density is sufficiently high to enable to plasmid DNA to participate in anion exchange with the ion exchange ligands of the matrix at a first pH. Once adsorbed to the matrix to form a complex, the complex can be washed in a wash solution with buffer and salt solution conditions designed to ensure the plasmid DNA remains adsorbed to the matrix throughout any such washing steps, while removing at least one contaminant. Finally, the plasmid DNA is eluted from the complex by combining the complex with an elution buffer having a second pH above that of the lysate and wash solutions, wherein the second pH is sufficiently high to promote desorption of the plasmid DNA from the matrix.

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The materials and methods of the present invention can be used to isolate genomic DNA from living tissue, including but not limited to blood, semen, vaginal cells, hair, buccal tissue, saliva, tissue culture cells, plant cells, placental cells, or fetal cells present in amniotic fluid and mixtures of body fluids. When the target nucleic acid is genomic DNA, it is necessary to disrupt the tissue to release the target genomic DNA from association with other material in the tissue, so the target genomic DNA can adhere to the pH dependent ion exchange matrix in the presence of a solution at the first pH. The resulting complex of matrix and genomic DNA is separated from the disrupted tissue, and washed to remove additional contaminants (if necessary). The genomic DNA is then eluted from the complex by combining the complex with an elution solution having a second pH which is higher than the first pH.

The following, non-limiting examples teach various embodiments of the invention. In the examples, and elsewhere in the specification and claims, volumes and concentrations are at room temperature unless specified otherwise. The magnetic silica particles used in the examples below were all either porous or nonporous MagneSil™ particles having the general preferred dimensions and siliceous oxide coating described as preferred above. More specifically, the porous MagneSil™ Particles used in the Examples below were taken from either of two batches of particles having the following characteristics: (1) a surface area of 55 m²/g, pore volume of 0.181 ml/g for particles of <600 Å diameter, pore volume of 0.163 ml/g for particles of >600 Å diameter, median particle size of 5.3µm, and iron leach of 2.8 ppm when assayed as described herein above using ICP; or (2) a surface area of 49 m²/g, pore volume of 0.160 ml/g (<600 Å diameter), pore volume of 0.163 ml/g (>600 Å diameter), median particle size of 5.5µm, and iron leach of 2.0 ppm.

One skilled in the art of the present invention will be able to use the teachings of the present disclosure to select and use magnetic particles other than the silica-based magnetic particles and ion exchange magnetic particles used to illustrate the methods and kits of the invention in the Examples, below.

The Examples should not be construed as limiting the scope of the present invention. Other magnetic silica particles and their use in the present method to concentrate cells, to clear solutions of disrupted biological material, and to isolate target nucleic acids

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from disrupted biological material will be apparent to those skilled in the art of chromatographic separations and molecular biology.

EXAMPLES

5 The following examples are given to illustrate various aspects of the invention, without limiting the scope thereof:

EXAMPLE 1 - GEL ELECTROPHORESIS

10 Samples of target nucleic acids isolated according to procedures described in Examples below were analyzed for contamination with non-target nucleic acids, and for size as follows. The samples were fractionated on an agarose gel of appropriate density (e.g., a 1.0% agarose gel was used to analyze plasmid DNA, while a 1.5% agarose gel was used to analyze RNA). The fractionated nucleic acid was visualized using a fluorescent label or by dyeing the gel with a DNA sensitive stain, such as ethidium
15 bromide or silver staining. The resulting fractionated, visualized nucleic acid was either photographed or visualized using a fluorimager and the resulting image printed out using a laser printer.

 In some cases, size standards were fractionated on the same gel as the target nucleic acid, and used to determine the approximate size of the target nucleic acid. In
20 every case where a gel assay was done, the photograph or fluorimage of the fractionated nucleic acid was inspected for contamination by non-target nucleic acids. For example, images of fractionated samples of plasmid DNA were inspected for RNA, which runs considerably faster than DNA on the same gel, and for chromosomal DNA, which runs considerably slower than plasmid DNA on the same gel. Images of isolated plasmid
25 DNA were also inspected to determine whether most of the plasmid DNA shown in the image is intact, supercoiled plasmid DNA.

EXAMPLE 2 - ABSORPTION SPECTROPHOTOMETRY

30 Samples of target nucleic acids isolated from various media, as described below, were also analyzed using absorption spectrophotometry. Absorption measurements were taken at wavelengths of 260, 280, and 230 nanometers (nm). A_{260}/A_{280} absorption ratios were computed from the measurements. An A_{260}/A_{280} of greater than or equal to 1.80

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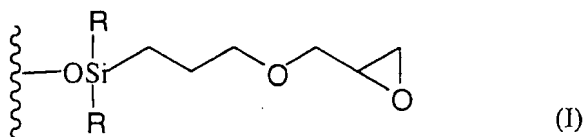
was interpreted to indicate the sample analyzed therein was relatively free of protein contamination. The concentration of nucleic acid in each sample was determined from the absorption reading at 260 nm (A_{260}).

5 EXAMPLE 3 - SYNTHESIS OF GLYCIDYL-HISTIDINE AND GLYCIDYL-ALANINE SILICA MAGNETIC ION EXCHANGE PARTICLES

Various two different pH dependent ion exchange ligands, glycidyl-histidine and glycidyl-alanine, were attached to porous silica magnetic particles according to the following procedure. The silica magnetic pH dependent ion exchange particles synthesized as
10 described herein were used to concentrate cells, clear lysates, or isolate target nucleic acids, as described in subsequent Examples, below.

A. Preparation of Glycidyl Modified Silica Magnetic Particles

1. Silica magnetic particles were activated by heating under vacuum at 110°C
15 overnight.
2. 10 g of the activated particles were suspended in 100 ml of toluene in a flask, and 3.2 ml of 3-glycidylpropyl-trimethoxysilane was added thereto.
3. The flask containing the mixture was fitted with a condenser and the reaction was refluxed for 5 hr. After cooling to room temperature, the reaction mixture sat for 48 hr at
20 room temperature.
4. The reaction mixture was then filtered and the retentate, including glycidyl-modified silica magnetic particles produced in the reflux reaction, were washed with toluene (2 x 100 ml), hexanes (2 x 100 ml) and ethyl ether (1 x 150 ml). The washed product was then left to dry in the air.
- 25 5. A small portion of the product was further dried in a 110°C oven and submitted for elemental analysis. The results (%C 0.75; %H 0.58) are consistent with glycidyl modification of silica gel particles, as illustrated in Formula (I), below. The wavy line in this and other formulae depicted herein and in the remaining Examples below represents the surface of a solid phase, a porous silica magnetic particle in this particular Example.



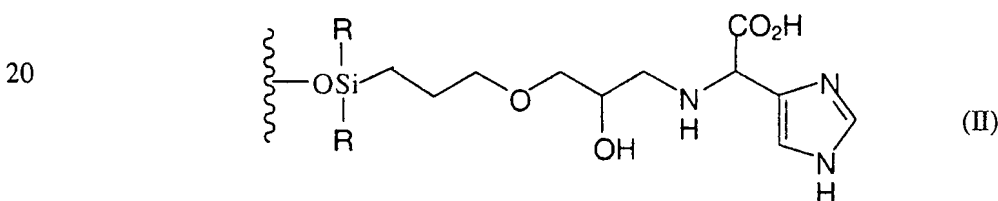
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wherein, R is $-\text{OH}$, OCH_3 , or $-\text{OCH}_2\text{CH}_3$.

6. The glycidyl-modified silica magnetic particles produced as described above were then further modified by the linkage of an amino acid, such as histidine, alanine, or cysteine to the particles, by reaction with the terminal ring of the glycidyl moiety, as described below.

B. Synthesis of Glycidyl-Histidine Modified Silica Magnetic Particles

1. 2.0 g. of D,L-histidine was dissolved in a mixture of 20 ml of tetrahydrofuran and 20 ml of water by heating the solution to reflux.
2. To this solution, 2 g of glycidyl-modified silica magnetic particles was added and the resulting suspension was refluxed overnight (18 hr).
3. After cooling to room temperature the reaction mixture was filtered, and the retentate, which included glycidyl-histidine modified silica magnetic particles, was washed once with 100 ml of acetone, three times with 150 ml of water, and once with 150 ml of ether. The solid was air dried.
4. A small portion of the dried solid from step 3 was further dried at 110°C and submitted for elemental analysis. Results: %C 1.35; %H 0.68; %N 0.50. This results are consistent with glycidyl-histidine linkage, such as is as shown in Figure (II), below:



wherein, R is $-\text{OH}$, OCH_3 , or $-\text{OCH}_2\text{CH}_3$.

25

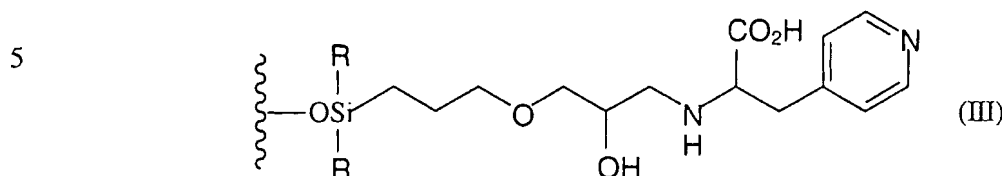
C. Synthesis of Glycidyl -Alanine Modified Silica Magnetic Particles

1. 3-(3-pyridyl)-D-alanine (1g) was dissolved in 20 ml of water.
2. To this solution 2 g. of glycidyl-modified silica magnetic particles were added, and the resulting mixture was refluxed overnight.
3. After cooling, the reaction mixture was filtered and washed twice with water, and once with ethyl ether.

30

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4. Elemental analysis of a sample of the product from step 3 showed: %C 0.98; %H 0.56; %N 0.20. This result is consistent with glycidyl-alanine modification, as illustrated in Formula (III), below:



wherein, R is -OH, OCH₃, or -OCH₂CH₃.

10

EXAMPLE 4 - PREPARATION OF A LYSATE OF PLASMID DNA

E. coli bacteria cells, DH5 α strain, were transformed with pGL3-Control Vector (Promega) plasmid DNA, grown overnight Luria Broth ("LB") medium at 37°C, then harvested by centrifugation.

- 15 The following solutions were used to prepare a lysate of the harvested cells, as described below:

Cell Resuspension Solution:

50mM Tris-HCl, pH 7.5

10mM EDTA

- 20 100 μ g/ml DNase-free ribonuclease A (RNase A)

Wizard[®] Neutralization Buffer (Promega Corp.):

1.32M KOAc (potassium acetate), pH 4.8

Cell Lysis Solution:

0.2M NaOH

- 25 1% SDS (sodium dodecyl sulfate)

A lysate of the transformed cells was produced as follows:

1. The cells from 1 to 10ml of bacteria culture were harvested by centrifuging the culture for 1-2 minutes at top speed in a microcentrifuge. The harvested cells were resuspended in 250 μ l of Cell Resuspension Solution, and transferred to a microcentrifuge tube. The resulting solution of resuspended cells was cloudy.
- 30

- 20 -

2. 250 μ l of Cell Lysis Solution was then added to the solution of resuspended cells and mixed by inversion until the solution became relatively clear, indicating the resuspended cells had lysed.

3. 350 μ l of Wizard[®] Neutralization Buffer was added to the lysate solution, and
5 mixed by inversion. The lysate became cloudy after the Neutralization Solution was added.

Each sample of lysate prepared as described above was cleared, either by centrifugation (control samples), or by using silica magnetic particles or silica magnetic ion exchange particles (test samples), as described in the Examples below.

10 EXAMPLE 5 LYSATE CLEARANCE BY CENTRIFUGATION OR SILICA MAGNETIC PARTICLES, FOLLOWED BY PLASMID DNA ISOLATION USING GLYCIDYL-HISTIDINE OR GLYCIDYL-ALANINE SILICA MAGNETIC PARTICLES

A. Preparation of Cleared Lysates

Four samples of lysates of 1ml cultures of DH5 α (pGL3) were prepared as described
15 in Example 4, above, except that 24 hour cultures in Circlegrow medium were used instead of overnight LB. Two of the samples were cleared by centrifugation. The other two samples were cleared by mixing the lysate with 150 μ l of silica magnetic particles (100mg/ml), vortexing the resulting mixture until debris in the lysate has adsorbed to the particles, and separating the silica magnetic particles from the solution by magnetic force,
20 using a magnetic separator.

B. Isolation of Plasmid DNA from Cleared Lysates

Plasmid DNA was then isolated from the samples of cleared lysate, as follows:

1. The cleared lysate solutions from both sets of samples were transferred to clean tubes containing 150 μ l of either glycidyl-histidine silica magnetic ion exchange particles
25 (hereinafter, "Mag-IE-glycidyl-histidine" particles) or glycidyl-alanine silica magnetic ion exchange particles (hereinafter, "Mag-IE-glycidyl-alanine particles"). and mixed by vortexing. The Mag-IE-glycidyl-alanine and Mag-IE-glycidyl-histidine particles were produced as described in Example 3, above.
2. After waiting 5 minutes for DNA binding to the particles, the solutions were placed
30 on a magnetic rack, allowed to sit for 2 minutes, and the solutions removed.

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3. The particles were then resuspended in 1.0 ml of nanopure water, the tubes inverted to wash the side-walls and cap, and placed back into a magnetic separator, which was inverted to wash the tube cap to removed suspended particles.
4. Step 3 (a water wash) was repeated 3 times, for a total of four washes.
5. The solution was removed from the tubes, and the DNA was eluted using (1) 10mM Tris HCl pH 8.5 for Mag-IE-glycidyl-histidine or (2) 20mM Tris HCl pH 9.5 for Mag-IE-glycidyl-alanine.

C. Assay of Results

A spectrophotometric assay was conducted on each eluent sample, as described in Example 2. Spectrophotometric results from the Mag-IE-glycidyl-histidine particle eluent showed a yield of 26 μ g of DNA and a high purity, with an A_{260}/A_{280} ratio of 1.85. Assay results from the Mag-IE-glycidyl-alanine particle eluent showed a yield of 25 μ g of DNA and a A_{260}/A_{280} ratio of 1.90, indicating a comparable purity to the eluent from the other species of IE particle described above.

All the eluents produced as described above were also assayed by gel electrophoresis, as described in Example 1, above. Intact plasmid DNA was detected in each sample, with no evidence of degradation or RNA contamination in any of the samples.

EXAMPLE 6 LYSATE CLEARANCE WITH SILICA MAGNETIC PARTICLES OR VARYING AMOUNTS OF MAG-IE-GLYCIDYL-HISTIDINE PARTICLES

The assay described below was performed to determine whether small quantities of silica magnetic ion exchange particles could clear lysate with sufficient efficiency that one could isolate intact plasmid DNA therefrom, which is substantially free of contaminants. Lysate cleared with 4mg of silica magnetic particles was used as a control. Plasmid DNA was isolated from both the control and test samples of cleared lysate, using Mag-IE-glycidyl-histidine, according to the same procedure, set forth below.

A. Lysate Clearing

Silica magnetic particles and varying amounts of Mag-IE-glycidyl-histidine particles were used, as follows, to prepare a cleared lysate. All the steps below were conducted in 1.5ml tubes, and at room temperature.

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1. A pellet of cells harvested, by centrifugation of a 50 ml overnight culture of DH5 α *E. coli* bacteria transformed with pGEM-3Zf⁺ plasmid DNA, were resuspended in 2.5 ml of Wizard[®] Resuspension Solution.
2. 265 μ l of the resuspended cells was added to each of eight tubes.
- 5 3. 250 μ l Wizard[™] Lysis solution was added to each tube of resuspended cells, and mixed gently, to avoid possible sheering of genomic DNA.
4. 350 μ l Wizard Neutralization solution was added to each tube of lysed cells, and mixed gently and thoroughly.
5. Mag-IE-histidine particles (100 mg/ml) were added to six of the samples from step 4, as follows: 10 μ l or 20 μ l or 40 μ l per lysate tube (in duplicate). 40 μ l of silica magnetic
10 particles (100mg/ml) were added to each of the remaining two samples. All the samples were mixed thoroughly, by vortexing.
6. The resulting particle/cell debris complex was separated from the lysate within each tube, using a magnetic separator. The caps of the tubes were washed four times, by inversion
15 of each tube. The tubes allowed to sit for 1 minute.

B. DNA Isolation

DNA was isolated from each of the cleared lysate samples, above, as described below:

- 20 1. Each cleared lysate solution sample, above, was transferred to a clean 1.5 ml tube containing 150 μ l of Mag-IE-glycidyl-histidine (100mg/ml), vortexed, and allowed to sit 5 minutes.
2. The resulting Mag-IE-glycidyl-histidine/DNA complex was then separated from the solution within each tube, using a magnetic separator. The tube caps were each washed four
25 times, by inversion. The tubes were allowed to sit for 1 minute.
3. The liquid was removed from each tube, and discarded.
4. The particles were washed with nanopure water, as follows. 1.0 ml nanopure water was added to each tube, and the particles resuspended therein. The Mag-IE-glycidyl-histidine particles were separated from the solution within each tube, using a magnetic
30 separator. The tube caps were each washed four times, by inversion. The tubes were allowed to sit for 1 minute. The liquid was removed from each tube and cap, and discarded,

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using the magnetic separator to retain the particles in each tube while the wash solution was discarded.

5. Step 4 was repeated twice, for a total of 3 washes.

6. Added 100 μ l 10mM Tris HCl, pH 8.0, to each tube, and resuspend the particles
5 contained therein by vortexing

7. The plasmid DNA was magnetically separated from the particles from the resulting eluent solution in each tube, and transferred to a clean tube.

C. Assay of Results

10 Each of the eluent samples produced as described above was assayed spectrophotometrically, as described in Example 2. The assay results are summarized in Table 2, below:

TABLE 1

PARTICLES & AMOUNT	A_{260}/A_{280}	NUCLEIC ACID YIELD
1mg of Mag-IE-glycidyl-histidine	1.73	37 μ g
	1.73	43 μ g
2 mg of Mag-IE-glycidyl-histidine	1.75	36 μ g
	1.76	38 μ g
4mg of Mag-IE-glycidyl-histidine	1.76	40 μ g
	1.76	38 μ g
4mg of Magnesil TM	1.80	36 μ g
	1.80	37 μ g

15 The samples assayed by spectrophotometric analysis, as described above, were also analyzed by gel electrophoresis, as described in example 1. Figure 1 shows a photograph of samples of each of the eluents, above, after being fractionated by gel electrophoresis and stained with ethidium bromide. The samples were loaded on the gel, from left to right, in the same order shown in Table 1, above. None of the samples showed any visible RNA, and the
20 intensity of the plasmid DNA bands is consistent with the yield data obtained by absorption spectrophotometry (as described in example 2).

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EXAMPLE 7: LYSATE CLEARANCE BY CENTRIFUGATION VS. USING SILICA MAGNETIC PARTICLES, FOLLOWED BY ISOLATION OF PLASMID DNA FROM CLEARED LYSATE USING SILICA MAGNETIC PARTICLES

In the following assay, centrifugation or silica magnetic particles were used to clear cell lysates of varying volumes of overnight cultures of the same transformants. Plasmid DNA was then isolated from each cleared lysate solution, using silica magnetic particles, and tested as described below.

A. Lysate Clearing

1. An overnight culture of DH5 α (pGL3) was centrifuged to obtain, in six replicates, 1.0 ml, 2.0ml, and 3ml cell pellets in 1.5 ml tubes. To each tube, 250 μ l of Resuspension Buffer was added, and the cells resuspended by vortexing.
2. 250 μ l of Wizard Lysis solution was added per tube, and gently mixed to avoid sheering genomic DNA.
3. 350 μ l Wizard Neutralization solution was added per tube, mixed gently and thoroughly.
4. To one set of triplicate samples, the tubes were centrifuged for 10 minutes at 12,000Xg to clear the lysate debris. The cleared supernatants were transferred to clean 1.5ml tubes and processed as described in section B, below.
5. To the other set of triplicate samples (3 of 1.0 ml, 3 of 2ml, 3 of 3ml), 50 μ l of resuspended silica magnetic particles (100 mg/ml) were added per lysate tube, and vortexed thoroughly.
6. The resulting particles/cell debris complex was separated from the solution in the tube, in a magnetic separator. Tube caps were washed by tube inversion (4X). Tubes were allowed to sit for 1 minute. The resulting cleared lysate was transferred from the each tube and processed as described in section B, below.

B. Isolation of DNA from Cleared Lysates

1. The cleared solutions from steps 4 and 6, above, were each placed in a clean 1.5ml tube containing 200 μ l of 5.0M guanidine thiocyanate, and vortexed. 150 μ l of silica magnetic particles (15 mg) was added per tube, vortexed, and allowed to sit 10 minutes.

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2. The resulting silica magnetic particle/ DNA complex was separated from the solution in the tube, on a magnetic separator. Tube caps were washed four times, by tube inversion, and allowed to sit in the separator for 1 minute.
3. Liquid was removed from each tube, including caps, and discarded.
- 5 4. Each tube was washed with 1 ml of 60mM KOAc / 10mM Tris-HCl (pH 7.5 at 25°C) / 60% ethanol, using vortexing to resuspend the particles.
5. The silica magnetic particle/DNA complex was separated from the wash solution in the tube, using a magnetic separator. Tube caps were washed four times, by tube inversion, and allowed to sit in the separator for 1 minute.
- 10 6. Liquid was removed from tube and caps, and discarded.
7. Steps 4-6 were repeated, for a total of 2 washes.
8. The tubes were allowed to air dry for 30 minutes to remove residual ethanol.
9. 100 μ l of nanopure water was added per tube, and the particles were resuspended thoroughly by vortexing. After 10 minutes at ambient temperature, the tubes were placed in
- 15 a magnetic separator, and the resulting eluent was transferred to clean 1.5ml tubes.

C. Analysis of Results

The eluent from each sample was analyzed with an absorption spectrophotometer at 230, 260, and 280 nm, as described in Example 3, above. The average value of test results
 20 obtained from each set of three samples of eluent, prepared as described above, is set forth in Table 2, below:

TABLE 2

VOL. CULTURE, CLERANCE MEANS	A₂₃₀	A₂₆₀	A₂₈₀	A₂₆₀/A₂₈₀	YIELD (μg DNA)
1 ml & Centrifugation	0.160	0.072	0.039	1.83	7.17
1 ml & Silica Magnetic Particles	0.176	0.094	0.053	1.77	9.36
2 ml & Centrifugation	0.197	0.121	0.667	1.82	12.0
2ml & Silica Magnetic Particles	0.189	0.103	0.058	1.79	10.3
3ml & Centrifugation	0.495	0.149	0.082	1.82	14.9

The results shown in Table 2, above, indicate comparable amounts of plasmid DNA were
 25 isolated from the same volumes of lysate cleared either by centrifugation or by silica magnetic particles. The A₂₃₀ and A₂₆₀/A₂₈₀ measurements from samples isolated from the

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same volumes of cultures lysed and cleared with each of the two means described above, indicates that both methods of isolation produced isolated DNA which appears to be free from contamination with low molecular weight alcohol or proteins.

Each of the samples of plasmid DNA isolated as described above was also assayed
5 by agarose gel electrophoresis, as described in Example 1. Initially, the agarose gel with the above samples loaded thereon was run only for a sufficient period of time for the plasmid DNA to migrate into the gel and become separated from any RNA present in each sample. Figure 2 is a photograph of the gel taken under UV light at this initial stage, after staining the gel with ethidium bromide. No sign of RNA contamination was apparent in any of the
10 lanes of the gel shown in Figure 2. The same gel was then electrophoresed for an additional period of time, to enable the plasmid DNA to become separated from any chromosomal DNA in each sample loaded thereon. Figure 3 is a photograph of the same gel, taken under the same conditions described above, after the gel had been run for a longer period of time. No sign of contamination with chromosomal DNA was apparent in any of the lanes of the
15 gel, in Figure 3.

EXAMPLE 8: CONCENTRATION OF CELLS, LYSATE CLEARING, AND DNA ISOLATION USING MAG-IE-GLYCIDYL-HISTIDINE PARTICLES

Mag-IE-glycidyl-histidine particles were used to concentrate cells prior to lysis, to
20 clear the lysate once the concentrated cells were lysed, and to isolate DNA from the resulting cleared lysate, as follows:

A. Cell Concentration:

1. 50 μ l of Mag-IE-glycidyl-histidine suspension was aliquoted into each of two 1.5ml tubes.
- 25 2. 500 μ l of an overnight culture of DH5 α /pGem3Zf+ was aliquoted into the two tubes prepared in step 1. These two samples were processed to harvest the cells as described in steps 4-6, below.
3. 500 μ l of the same culture used in step 2 was also aliquoted into each of two empty 1.5ml centrifuge tubes, and spun in a centrifuge to harvest the cells. The supernatant was
30 discarded, and the harvested cells processed as described in section B, below.
4. 300 μ l of 5M NaCl was added to each tube of Mag-IE-glycidyl-histidine and overnight culture, and mixed thoroughly.

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5. 800 μ l of room temperature isopropanol was added to each tube, and mixed thoroughly for a final concentration of 94M NaCl / 50% IPA.
6. The resulting Mag-IE-glycidyl-histidine/cells complex was separated from the solution in each tube, in magnetic separator. The solution was discarded, and the harvested
5 cells processed as described in section B, below.

B. Lysate Clearing and DNA Isolation

1. 250 μ l Wizard Resuspension solution was added to both sets of tubes, the tubes with cells pelleted in a centrifuge and the tubes with cells complexed with Mag-IE-glycidyl-
10 histidine particles. In both cases, the solutions were mixed thoroughly until the cells were resuspended in each solution.
2. 250 μ l of Wizard Lysis solution was added to each tube, and gently mixed to avoid sheering genomic DNA
3. 350 μ l of Wizard neutralization solution was added, and mixed gently and
15 thoroughly.
4. The resulting Mag-IE-glycidyl-histidine/cell debris complex was separated from the lysate within each tube, using a magnetic separator.
5. Each resulting cleared lysate solution was transferred to a clean 1.5 ml tube containing 50 μ l Mag-IE-glycidyl-histidine, and incubated 2 minutes at room temperature, to
20 enable DNA to adhere to the particles.
6. The resulting Mag-IE-glycidyl-histidine/ DNA complex was separated from the solution in the tube, using a magnetic separator.
7. The liquid in the tube was removed and discarded.
8. Each tube was washed with 1.0 ml nanopure water, and the particles suspended
25 therein. The particles were separated from the water in each tube, using a magnetic separator. The liquid was removed and discarded.
9. Step 8 was repeated three times, for a total of four washes.
10. 100 μ l of 20mM Tris pH 9.5, an elution buffer, was then added to each tube. The particles were resuspended in the elution buffer.
- 30 11. Magnetic force was used to separated the Mag-IE-glycidyl-histidine particles from the resulting eluent solution.

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C. Assay Results

The four samples of DNA isolated from cells which were concentrated by either centrifugation or using Mag-IE-glycidyl-histidine particles, as described above, were assayed spectrophotometrically, as described in Example 1, above. The results of the spectrophotometric analysis are presented in Table 3, below:

TABLE 3

SAMPLE	A_{260}/A_{280}	YIELD
Centrifugation used to concentrate cells	1.84	7.8 μ g
	1.85	8.1 μ g
Mag-IE glycidyl-histidine used to concentrated cells	1.78	9.5 μ g
	1.80	8.2 μ g

10

EXAMPLE 9: CLEARING MOUSE TISSUE HOMOGENATES USING MAG-IE-GLYCIDYL-HISTIDINE, AND ISOLATING DNA AND RNA THEREFROM USING MAG-IE-GLYCIDYL-HISTIDINE

The following protocol was used to clear homogenates of frozen mouse liver, kidney, and spleen tissue, and to isolate RNA and DNA therefrom:

15

A. Homogenate Clearance

1. A sample of each tissue was homogenized in a solution of 4.5M guanidine thiocyanate (GTC)/ 132mM KOAc pH 4.8, wherein, for every 1mg of tissue, 1 μ l of homogenization solution was used. 120mg of liver, 320mg of kidney, and 142mg of spleen were homogenized.

20

2. The resulting homogenized mixture was diluted 7X with RNase free nanopure water for mouse liver, 6X RNase free nanopure water for kidney, and 12X RNase free nanopure water for spleen. After the addition of nanopure water (liver = 840 μ l, spleen = 1.7 ml, and kidney = 1.9 ml), each sample was vortexed.

25

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3. "1/2 X" volume of Mag-IE-glycidyl-histidine (100mg/ml) was added to each solution, and vortexed. The resulting mixture was then magnetically separated for 10 minutes.

5 B. Isolation of Nucleic Acids from Cleared Homogenate

1. An aliquot of each cleared solution separated from the Mag-IE-glycidyl-histidine particles, as described above, was transferred to a clean tube containing Mag-IE-glycidyl-histidine particles. For the liver and spleen samples, 100µl of cleared solution was added to 100µl of Magnesil-IE-glycidyl-histidine (100mg/ml), the mixture was vortexed, allowed to sit for 2 minutes, then allowed to sit in a magnetic separator for 2 minutes. For the kidney sample, 400µl of cleared solution was added to 1 ml of RNase free nanopure water, then 100µl of Magnesil-IE-glycidyl-histidine (100mg/ml) was added, the mixture was vortexed, allowed to sit for 2 minutes, then allowed to sit in a magnetic separator for 2 minutes.
2. The solution was then removed from each tube, and each tube was washed with 1.0 ml RNase free nanopure water, vortexed, and placed back in the magnetic separator. The tube cap was washed by inversion of the tubes in the magnetic rack. After 2 minutes, the wash solution was removed. This wash step was repeated two times, for a total of 3 washes.
3. The nucleic acids were eluted in 100µl of 10mM Tris HCl, pH 9.5.

20 C. Analysis of Results

The eluted DNA and RNA was visualized by gel electrophoresis (see example 1) as shown in figures 4, 5, and 6. Figure 4 shows a photograph of mouse liver DNA and RNA isolated as described above, fractionated by gel electrophoresis along with λ Hind III marker. Both DNA and RNA appear to be present in each eluent.

- 25 Figure 5 shows DNA and RNA isolated from mouse spleen and kidney as described above, after fractionation by gel electrophoresis. Samples were loaded on the gel as follows:

- 30
- Lane 1: λ Hind III marker
 - Lane 2: Spleen, 0µl removed
 - Lane 3: Spleen, 20µl removed
 - Lane 4: Spleen, 40µl removed

- 30 -

Lane 5: Spleen, all removed

Lane 6: λ Hind III marker

Lane 7: Kidney, 0 μ l removed

Lane 8: Kidney, 20 μ l removed

5 Lane 9: Kidney, all removed

Lane 10: λ Hind III marker

Figure 6 shows samples of mouse liver RNA and DNA isolated as described above, after digestion with DNase and fractionation by gel electrophoresis. Lanes 1 and 4 contain λ Hind III marker, while lanes 2 and 3 contain mouse liver nucleic acid isolated from 200 μ l and 400 μ l of homogenate, respectively, according to the procedure described above.

EXAMPLE 10: CONCENTRATION OF WHITE BLOOD CELLS, LYSATE CLEARING, AND DNA ISOLATION FROM WHOLE BLOOD USING MAG-IE-GYLCIDYL-HISTIDINE PARTICLES, NON-POROUS MAGNESIL-IE-GLY-HISTIDINE PARTICLES, AND MAGNESIL™ PARTICLES USING HUMAN WHOLE BLOOD

Mag-IE-glycidyl-histidine particles, Non-Porous Mag-IE-glycidyl-histidine particles and Magnesil™ particles were used to either (a) concentrate white blood cells, clear the lysate once the concentrated cells were lysed, and to isolate DNA from the resulting cleared lysate, or (b) clear the lysate produced from centrifugal concentrated white blood cells, clear the lysate, and to isolate DNA from the resulting cleared lysate.

A. Use of Mag-IE-glycidyl-histidine Particles With Ion Exchange Wash:

Magnetic clearing of blood lysate and purification of genomic DNA using solutions from Promega's Wizard Genomic DNA Purification kit (see, Promega's Technical Manual #TM50), and Mag-IE-glycidyl-histidine particles: All steps were at room temperature. Mag-IE-glycidyl-histidine particles were used with an ion exchange wash to concentrate white blood cells, to clear a lysate of the cells, and to isolate genomic DNA therefrom, as follows:

1. 1.0 ml of blood was placed in a 15 ml tube containing 3.0 ml of Wizard Genomic Cell Lysis solution, mixed, and incubated for 10 minutes.
2. 1.0 ml of 5.0 M NaCl was added, and mixed.

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3. 50µl of Mag-IE-glycidyl-histidine particles in a 100mg/ml solution was added to the tube, and mixed.
4. 5.0 ml of isopropanol was added and mixed, and incubated for 2 minutes, then placed on a magnetic rack for 5 minutes.
- 5 5. The solution was removed and discarded.
6. The tubes were removed from the magnetic rack and vortexed for 5 seconds.
7. 1.0 ml of Nuclei Lysis solution was added, the tube was vortexed for 5 seconds, and incubated for 5 minutes.
8. 330 µl of Wizard Genomic Protein Precipitation solution was added, the tube was
- 10 vortexed for 5 seconds, and the tube was placed on a magnetic rack for 5 minutes.
9. The cleared lysate solution was removed from the first tube and placed into a second tube containing 200 µl of Mag-IE-glycidyl-histidine particles (100 mg/ml). and mixed.
10. 0.5 ml of 0.5 M sodium citrate, pH 5.0 (pH adjusted to 5.0 with citric acid) was added, and the solution mixed. 8.0 ml of nanopure water was added, the solution mixed, the
- 15 tube was incubated for 1 minute, and placed on a magnetic rack for 2 minutes.
11. The solution was removed and discarded.
12. 5.0 ml of 66 mM potassium acetate, pH 4.8 (pH adjusted with acetic acid) was added, the tube vortexed for 5 seconds, and the tube placed on a magnetic rack for 2 minutes.
- 20 13. The solution was removed and discarded, and 2.0 ml of 66 mM potassium acetate / 600 mM NaCl , pH 4.8 was added, the tube mixed, and placed on a magnetic rack for 2 minutes.
14. The solution was removed and discarded.
15. 2.0 ml of 66 mM potassium acetate, pH 4.8, 450 mM NaCl was added, the tube was
- 25 vortexed for 5 seconds, and the tube placed on a magnetic rack for 2 minutes.
16. The solution was removed and discarded.
17. 10 ml of nanopure water was added, mixed, and the tube placed onto a magnetic rack for 2 minutes, after which time the solution was discarded.
18. Step 17 was repeated twice, for a total of 3 X 10ml nanopure water washes.
- 30 19. After removal from the magnetic rack, DNA was eluted in 400 µl of 90mM Tris HCl, pH 9.5 for 5 minutes. The tube was then placed on a magnetic rack for 5 minutes.

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Mag-IE-glycidyl-histidine particles were also used to clear a lysate of white blood cells isolated by centrifugation, before isolating genomic DNA therefrom using the same particles. The same procedure described above was used, except that Steps 2-4 were replaced by centrifugation for 10 minutes at 800 X g, followed by removal of the lysed red blood cell debris, and vortexing the cell pellet to resuspend the white blood cells. Also, in step 8, 50 µl of Mag-IE-glycidyl-histidine particles were added after the vortexing step, and followed by five seconds of vortexing, prior to placement of the tube into the magnetic rack.

10 B. MagneSil™ Particles and Guanidine Thiocyanate

Magnetic clearing of blood lysate and purification of genomic DNA using solutions from Promega's Wizard® Genomic DNA Purification kit, and MagneSil™ particles: guanidine thiocyanate protocol, as described below. All steps were at room temperature

1. 1.0 ml of blood was placed in a 15 ml tube containing 3.0 ml of Wizard Genomic Cell Lysis solution, mixed, and incubated for 10 minutes.
2. 1.0 ml of 5.0 M NaCl was added, and mixed.
3. 50 µl of MagneSil™ Particles, (100 mg/ml) was added to the tube, and mixed.
4. 5.0 ml of isopropanol was added and mixed, and incubated for 2 minutes, then placed on a magnetic rack for 5 minutes.
5. The solution was removed and discarded.
6. The tubes were removed from the magnetic rack and vortexed for 5 seconds.
7. 1.0 ml of Nuclei Lysis solution was added, the tube was vortexed for 5 seconds, and incubated for 5 minutes.
8. 330 µl of Wizard Genomic Protein Precipitation solution was added, the tube was vortexed for 5 seconds, and the tube was placed on a magnetic rack for 5 minutes.
9. 200 µl of MagneSil™ Particles (100 mg/ml) was added to a clean tube, placed on a magnetic rack for 1 minute, and the solution removed. To this tube, the cleared lysate solution was added from the tube in step 8, and mixed.
10. 2.0 ml of 5 M guanidine thiocyanate (GTC) was added, the tube mixed, incubated 2 minutes, and placed on a magnetic rack for 5 minutes.
11. The solution was removed and discarded.

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12. 5.0 ml of SV Total RNA Column Wash was added, the tube was vortexed for 5 seconds, and the tube placed on a magnetic rack for 2 minutes.
 13. The solution was removed and discarded
 14. Steps 12-13 were repeated, for a total of 2 washes.
 - 5 15. 5.0 ml of 80% ethanol was added, and the tube vortexed for 5 seconds, and the tube placed on a magnetic rack for 2 minutes.
 16. The solution was removed and discarded.
 17. Steps 15-16 were repeated 2 times, for a total of 3 washes.
 18. The tubes were air-dried for 60 minutes in the magnetic rack.
 - 10 19. After removal from the magnetic rack, DNA was eluted in 400 μ l of Wizard Genomic Renaturation Solution for 5 minutes. The tube was then placed on a magnetic rack for 5 minutes.
 20. The DNA containing solution was removed to a clean tube.
- 15 For the isolation of white blood cells by centrifugation, followed by clearing of the lysate and isolation of DNA with MagneSil™ Particles: Steps 2-4 were replaced by centrifugation for 10 minutes at 800 X g, followed by removal of the lysed red blood cell debris, and vortexing the cell pellet to resuspend the white blood cells. Additionally, 50 μ l of MagneSil™ particles were added in step 8 after the vortexing step, and followed by five
- 20 seconds of vortexing, prior to placement of the tube into the magnetic rack.

C. Non-Porous-Mag-IE-glycidyl-histidine Particles and Isopropanol

- Magnetic clearing of blood lysate and purification of genomic DNA using solutions from Promega's Wizard Genomic DNA Purification kit, and Non-porous MagneSil-IE-glycidyl-histidine particles, as follows. All steps were at room temperature
- 25
1. 1.0 ml of blood was placed in a 15 ml tube containing 3.0 ml of Wizard Genomic Cell Lysis solution, mixed, and incubated for 10 minutes.
 2. 1.0 ml of 5.0M NaCl was added, and mixed.
 3. 100 μ l of Non-Porous-Mag-IE-glycidyl-histidine in a solution of 100mg/ml was
 - 30 added to the tube, and mixed.
 4. 5.0 ml of isopropanol was added and mixed, and incubated for 2 minutes, then placed on a magnetic rack for 5 minutes.

- 34 -

5. The solution was removed and discarded.
6. The tubes were removed from the magnetic rack and vortexed for 5 seconds.
7. 1.0 ml of Nuclei Lysis solution was added, the tube was vortexed for 5 seconds, and incubated for 5 minutes.
- 5 8. 330 μ l of Wizard Genomic Protein Precipitation solution was added, the tube was vortexed for 5 seconds, and the tube was placed on a magnetic rack for 5 minutes.
9. The cleared lysate solution was removed from the first tube and placed into a second tube containing 20 mg of Non-Porous Mag-IE-glycidyl-histidine (200 μ l of 100 mg/ml, placed on a magnetic rack and the solution removed), and mixed.
- 10 10. 1.0 ml of isopropanol was added, the solution mixed, incubated 2 minutes, then placed in a magnetic rack for 2 minutes.
11. The solution was removed and discarded.
12. 2.0 ml of 66 mM potassium acetate, pH 4.8 (pH adjusted with acetic acid) was added, and the tube vortexed 5 seconds, incubated 1 minute, and the tube placed on a magnetic rack for 2 minutes.
- 15 13. The solution was removed and discarded.
14. 2.0 ml of nanopure water was added, mixed, and the tube placed onto a magnetic rack for 2 minutes, after which time the solution was discarded.
15. Step 18 was repeated twice, for a total of 3 X 2 ml nanopure water washes.
- 20 16. After removal from the magnetic rack, DNA was eluted in 400 μ l of 90mM Tris HCl, pH 9.5 for 5 minutes. The tube was then placed on a magnetic rack for 5 minutes.
17. The DNA containing solution was removed to a clean tube.

For the isolation of white blood cells by centrifugation, followed by clearing of the lysate and isolation of DNA with Non Porous-Mag-IE-glycidyl-histidine: Steps 2-4 were replaced by centrifugation for 10 minutes at 800 X g, followed by removal of the lysed red blood cell debris, and vortexing the cell pellet to resuspend the white blood cells. Additionally, 100 μ l of NP-Mag-IE-glycidyl-histidine particles were added in step 8 after the vortexing step, and followed by five seconds of vortexing, prior to placement of the tube into the magnetic rack.

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D. MagneSil-IE-glycidyl-histidine and isopropanol

The "Non-Porous-Mag-IE-glycidyl-histidine and Isopropanol" method described above was also used with porous Mag-IE-glycidyl-histidine particles. The only changes in the protocol were the use of 50 μ l of Mag-IE-glycidyl-histidine instead of 100 μ l of Non-Porous-Mag-IE-glycidyl-histidine particles in step 3, and the use of porous Mag-IE-glycidyl-histidine particles in step 8.

For the isolation of white blood cells by centrifugation, followed by clearing of the lysate and isolation of DNA with Mag-IE-glycidyl-histidine particles: Steps 2-4 were replaced by centrifugation for 10 minutes at 800 X g, followed by removal of the lysed red blood cell debris, and vortexing the cell pellet to resuspend the white blood cells. Additionally, 50 μ l of Mag-IE-glycidyl-histidine particles were added in step 8 after the vortexing step, and followed by five seconds of vortexing, prior to placement of the tube into the magnetic rack.

E. Assay Results

The A_{260}/A_{280} data and DNA yields were calculated from UV spectrophotometry, except for the porous Mag-IE-glycidyl-histidine particles white blood cell concentration samples, where estimates taken from gel electrophoresis were used, as denoted by "(gel)" below. These results are summarized in Table 4, below:

TABLE 4

PARTICLES USED	METHOD USED	A_{260}/A_{280}	YIELD (μ g)
Porous Mag-IE-glycidyl-histidine	Spin Cells, Clear Lysate (salt wash)	1.77	11
		1.79	7
Porous Mag-IE-glycidyl-histidine	Concentrate Cells with Particles,	1.27	10 (gel)
	Clear Lysate (salt wash)	1.29	8 (gel)
MagneSil™	Spin Cells, Clear Lysate (guanidine thiocyanate)	1.75	12
		1.82	10
MagneSil™	Concentrate with Particles, Clear Lysate (guanidine thiocyanate)	1.75	8
		1.71	7

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PARTICLES USED	METHOD USED	A_{260}/A_{280}	YIELD (μ g)
Porous Mag-IE-glycidyl-histidine	Spin Cells, Clear Lysate (isopropanol)	1.76	10
		1.78	15
Porous Mag-IE-glycidyl-histidine	Concentrate with Particles, Clear Lysate (isopropanol)	1.71	9
		1.75	13
Non-Porous Mag-IE-glycidyl-histidine	Spin Cells, Clear Lysate (isopropanol)	1.77	4
		1.78	5
Non-Porous Mag-IE-glycidyl-histidine	Concentrate with Particles, Clear Lysate (isopropanol)	1.65	5
		1.57	7

CLAIMS

We claim:

1. A method of using magnetic particles to concentrate or harvest cells, comprising the steps of:
 - 5 (a) combining a solution with cells contained therein with magnetic particles, under conditions wherein the cells form a complex with the magnetic particles; and
 - (b) isolating the complex from the solution by application of magnetic force.
2. The method of claim 1, wherein the solution with cells contained therein is growth
10 medium with a culture of bacteria suspended therein.
3. The method of claim 1, wherein the cells are blood cells.
4. The method of claim 3, wherein the cells are mammalian white blood cells and the
15 solution with cells contained therein is whole blood.
5. The method of claim 1, wherein the magnetic particles are silica magnetic particles.
6. The method of claim 1, wherein the magnetic particles are pH-dependent ion
20 exchange magnetic particles.
7. The method of claim 6, wherein the pH dependent ion exchange magnetic particles are selected from the group consisting of glycidyl-histidine modified silica magnetic particles, and glycidyl-alanine modified silica magnetic particles.
25
8. A method of clearing a solution of disrupted biological material, according to steps comprising:
 - (a) providing a solution comprising a disrupted biological material;
 - (b) combining the solution with second magnetic particles under conditions wherein
30 the disrupted biological material forms a complex with the second magnetic particles; and
 - (c) separating the complex from the solution by application of magnetic force.

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9. The method of claim 8, wherein the disrupted biological material is a bacterial cell lysate.
- 5 10. The method of claim 8, wherein the disrupted biological material is a homogenate of mammalian tissue.
11. The method of claim 8, wherein the disrupted biological material is a lysate of blood.
- 10 12. The method of claim 11, wherein the disrupted biological material is a lysate of mammalian white blood cells isolated from whole blood.
13. The method of claim 8, wherein the second magnetic particles are silica magnetic particles.
- 15 14. The method of claim 8, wherein the second magnetic particles are second pH dependent ion exchange magnetic particles.
15. The method of claim 14, wherein the second pH dependent ion exchange magnetic particles are selected from the group consisting of glycidyl-histidine modified silica magnetic particles, and glycidyl-alanine modified silica magnetic particles.
- 20 16. The method of claim 8, wherein the method further comprises producing the disrupted biological material provided in step (a), according to the steps comprising:
- 25 combining a solution with cells contained therein with first magnetic particles, under conditions wherein the cells form a complex with the first magnetic particles; isolating the complex from the solution by application of magnetic force; and disrupting the cells.
- 30 17. The method of claim 16, wherein the first magnetic particles are silica magnetic particles.

18. The method of claim 16, wherein the first magnetic particles are first pH-dependent ion exchange magnetic particles.

19. The method of claim 18, wherein the first pH-dependent ion exchange magnetic
5 particles are selected from the group consisting of glycidyl-histidine modified silica magnetic particles, and glycidyl-alanine modified silica magnetic particles.

20. The method of claim 16, wherein the first magnetic particles are the same as the second magnetic particles.

10

21. A method of isolating a target nucleic acid from a disrupted biological material, comprising the target nucleic acid, a first non-target material, and a second non-target material, comprising the steps of:

15 (a) combining a solution of the disrupted biological material with first magnetic particles under conditions wherein the first non-target material forms a first complex with the first magnetic particles;

(b) separating the first complex from the solution of disrupted biological material by application of magnetic force, forming a cleared solution comprising the target nucleic acid and the second non-target material;

20 (c) combining the cleared solution with second magnetic particles under conditions wherein the target nucleic acid adsorbs to the second magnetic particles, forming a second complex;

(d) isolating the second complex from the cleared solution;

25 (e) washing the second complex by combining the second complex with a wash solution and separating the second complex from the wash solution by magnetic force; and

(f) combining the washed second complex with an elution solution, under conditions wherein the target material is desorbed from the second magnetic particles.

30 22. The method of claim 21, wherein the disrupted biological material is selected from the group consisting of a lysate of bacteria cells, a lysate of blood cells, and a homogenate of tissue.

23. The method of claim 21, wherein the target nucleic acid is plasmid DNA.
24. The method of claim 21, wherein the target nucleic acid is genomic DNA.
- 5 25. The method of claim 21, wherein the target nucleic acid is RNA.
26. The method of claim 21, wherein the first magnetic particles are selected from the group consisting of: silica magnetic particles, and pH-dependent ion exchange magnetic
10 particles.
27. The method of claim 21, wherein the second magnetic particles are selected from the group consisting of: silica magnetic particles, and pH-dependent ion exchange magnetic
15 particles.
30. The method of claim 21, wherein the first non-target material comprises cell debris or homogenized tissue and a precipitate, wherein the precipitate is of material selected from the group consisting of proteins, non-target nucleic acids, and lipids.
- 20 31. The method of claim 21, wherein the second non-target material remains in solution when the target nucleic acid is adsorbed to the second magnetic particles in step (c).
32. A kit for isolating a target nucleic acid from a disrupted biological material, comprising:
25 a first container of first magnetic particles with the capacity to form a first complex with first non-target material in a first solution of disrupted biological material comprising the first non-target material and the target nucleic acid; and
a second container of second magnetic particles with the capacity to form a
second complex with the target nucleic acid, under solution conditions designed to
30 promote the specific adsorption of the target nucleic acid to the second magnetic particles.

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33. The kit of claim 32, wherein the first magnetic particles, the second magnetic particles, or both the first and second magnetic particles are silica magnetic particles.

5 34. The kit of claim 32, wherein the first magnetic particles, the second magnetic particles, or both the first and second magnetic particles are pH-dependent ion exchange particles.

35. The kit of claim 32, further comprising a wash solution, configured for use in washing the second complex prior to desorption of the target nucleic acid therefrom.

10

36. The kit of claim 33, further comprising an elution solution, configured to promote desorption of the target nucleic acid from the second complex.

MagIE-glycidyl-histidine| Magnasil™
1mg 2mg 4mg | 4mg



FIG. 1

Spin Mag Spin Mag Spin Mag
1ml 1ml 2ml 2ml 3ml 3ml

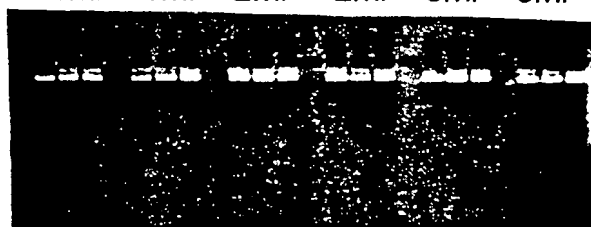
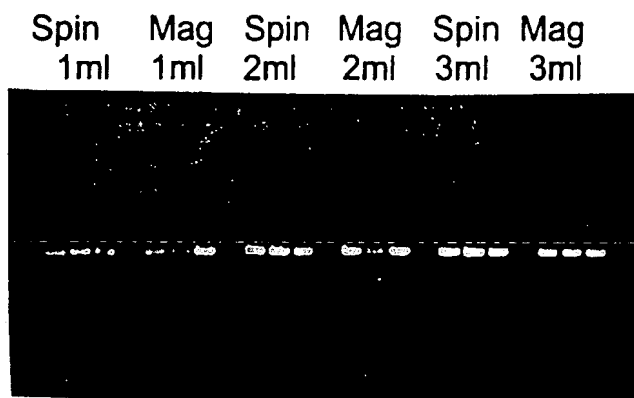


FIG. 2

**FIG. 3**

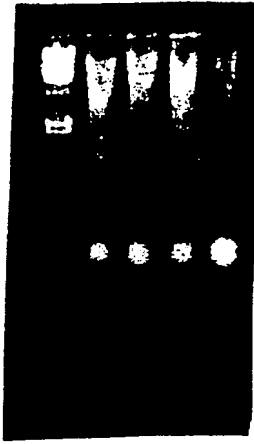
Marker	Samples			
λ Hind III	a	b	c	d
				

FIG. 4

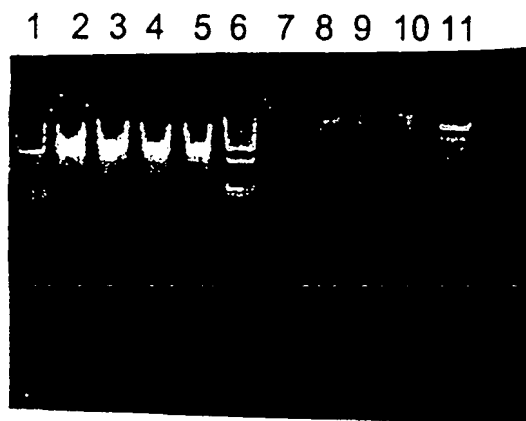


FIG. 5



FIG. 6

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 99/31207

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/10 C12N5/00 C12N1/02

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, BIOSIS, PAJ, MEDLINE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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X	DE 195 12 368 A (BOEHRINGER MANNHEIM GMBH) 2 October 1996 (1996-10-02) abstract page 5, line 36 - line 45 page 1, line 64 - page 2, line 1 -/-	1-4

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

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- *&* document member of the same patent family

Date of the actual completion of the international search

22 September 2000

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INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 99/31207

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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A	US 5 898 071 A (HAWKINS TREVOR) 27 April 1999 (1999-04-27) abstract column 2, line 20 - line 29 column 3, line 5 - line 11 column 3, line 40 column 4, line 59 - line 61 claims 1,6	8,13-15, 21, 23-27, 32-36

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